Review Chlorogenic acids and other cinnamates – nature, occurrence and dietary burden[†]

Michael N Clifford

Food Safety Research Group, School of Biological Sciences, University of Surrey, Guildford, Surrey, GU2 5XH, UK

Abstract: This review defines the range of forms in which cinnamates (*p*-coumarates, caffeates, ferulates and sinapates) occur in foods and beverages subdividing them into (i) the classic chlorogenic acids and close allies, (ii) other esters, amides and glycosides, and (iii) transformation products formed during processing. Cinnamate derivatives which would not release cinnamic acid by hydrolysis are excluded. The quantitative data are reviewed concisely and attention is drawn to certain shortcomings, in particular a complete absence of data for certain commodities (breakfast cereals, baked goods, tomato products and nuts) and minimal data for pulses, legumes and processed or cooked foods. In addition, more data are required for the edible portion of modern varieties. By extrapolating from such data as are available the important source(s) (i) of individual cinnamates (regardless of the conjugate type) and (ii) of each major class of conjugate, have been identified as follows:

- (i) Cinnamates: caffeic acid: coffee beverage, blueberries, apples, ciders; p-coumaric acid: spinach, sugar beet fibre, cereal brans; ferulic acid: coffee beverage, citrus juices, sugar beet fibre, cereal brans; sinapic acid: broccoli, kale, other leafy brassicas, citrus juices.
- (ii) Conjugates: caffeoylquinic acids: coffee beverage, blueberries, apples, ciders; p-coumaroylquinic acids: sweet cherries; feruloylquinic acids: coffee beverage; tartaric conjugates: spinach, lettuce, grapes and wines; malic conjugates: lettuce, spinach, possibly legumes; rosmarinic acid: culinary herbs, mixed herbs, possibly stuffings; cell wall conjugates: spinach, sugar beet fibre, cereal brans.

It seems likely that the UK population will fall into several categories depending on (i) their consumption of coffee, (ii) their consumption of bran, and (iii) their consumption of citrus. Those who drink several cups of coffee per day augmented by bran and citrus might easily ingest 500-800 mg *cinnamates* (or even 1 g for the greatest coffee ingest consumption) whereas those who eschew all these and take little fresh fruit or vegetables might struggle to consume 25 mg. (C) 1999 Society of Chemical Industry

Keywords: review; chlorogenic acid; cynarin; cinnamates; caffeic; ferulic; *p*-coumaric; sinapic; caftaric; coutaric; fertaric; chicoric; phaseolic; avenathramides; fruits; vegetables; berries; pome fruits; stone fruits; citrus; grapes; legumes; brassicas; herbs; cereals; beers; wines; tea; coffee; cocoa; cider; maté; bran; fibre; human diet; Apiaceae; Asteraceae; Boraginaceae; Chenopodiaceae; Fabaceae; Lamiaceae; Rosaceae; Solanaceae; Vitaceae

INTRODUCTION

Possibly the first report referring to chlorogenic acids was that published in 1837 by Robiquet and Boutron,¹ but the term itself does not seem to have been introduced until 1846 by Payen.^{2,3} In 1932 chlorogenic acid was shown to be a caffeic acid conjugate of quinic acid.⁴

Since then a vast literature has developed and it has become apparent that there are many cinnamoylquinic conjugates, that these are only one of many conjugated forms of the cinnamic acids, and that such conjugates are extremely widespread in the plant kingdom. It is not easy to summarise this literature and even less easy to put this information into the context of human food. Many of the data available relate to non-food sources although some plants studied may be close botanical relatives of plants consumed and others may be of interest as herbal remedies. Even when foodstuffs have been analysed, data are often qualitative, or for a very limited number of samples and varieties (not necessarily those of current horticultural or agricultural

Guildford, Surrey, GU2 5XH, UK

Received 15 June 1998; revised version received 7 September 1998; accepted 15 October 1998

t Based on a paper presented at Ferulate '98, IFR, Norwich, 8–11 July 1998.

^{*} Correspondence to: Michael N Clifford, Food Safety Research Group, School of Biological Sciences, University of Surrey,

importance) and quite often not solely for edible tissue. Only occasionally has consideration been given to maturity or season and rarely to the effects of processing or cooking.

It is not easy to identify unequivocally one component as a particular conjugate among many possible and structurally similar substances and it is now known that some early identifications were in error. For example, the original identification of cinnamoyl-glycerols in pineapple as 1,4-di-pCoQA,⁵ the incorrect identification of a caffeovltartaric acid glucoside in grapes and of 1,5-diCOA in coffee, corrected by Baranowski and Nagel⁶ and Clifford⁷ respectively, and the early confusion over the conjugates in cranberries discussed by Marwan and Nagel.⁸ There may be others as yet unrecognised, especially in view of some apparent inconsistencies in the data available.

THE NATURE OF THE CINNAMATE CONJUGATES The chlorogenic acids

Classically, chlorogenic acids (CGA) are a family of esters formed between certain *trans*-cinnamic acids and quinic acid (1L-1(OH),3,4/5-tetrahydroxycyclo-hexane carboxylic acid) which has axial hydroxyls on carbons 1 and 3, and equatorial hydroxyls on carbons 4 and 5. During processing, *trans* isomers may be partially converted to *cis*.

Using the preferred IUPAC numbering,⁹ the commonest individual chlorogenic acid is 5-O-caffeoylquinic acid (5-CQA). It is the only one commercially available and is still often called chlorogenic acid or 3-CQA (pre-IUPAC numbering) but the use of both terms should be discouraged. (Please note that in this review all citations will use the IUPAC system and if necessary numbering has been changed for consistency and clarity.)

These classic CGA may be subdivided by the identity, number and position of the acyl residues. The following subgroups are known:

- the relatively widespread mono-esters of caffeic acid, ie caffeoylquinic acids (CQA), *p*coumaroylquinic acids (*p*CoQA) and feruloyl quinic acids (FQA);
- di-esters, tri-esters and the single tetra-ester of caffeic acid, ie diCQA, triCQA and tetraCQA, the latter two groups being characteristic of Asteraceae;¹⁰⁻¹⁴
- mixed di-esters of caffeic and ferulic acid, ie caffeoylferuloylquinic acids (CFQA) which are characteristic of robusta coffee (*Coffea canephora*, Rubiaceae)¹⁵⁻¹⁸ or caffeic and sinapic acid, ie caffeoylsinapoylquinic acids (CSiQA) as found in *Gardenia jasminioides* (Rubiaceae);¹⁹
- mixed esters involving various permutations of between one and three residues of caffeic acid with one or two residues of a dibasic aliphatic acid (eg glutaric, oxalic, succinic) which are characteristic of Asteraceae.^{20–22}

It is convenient to extend this list of classic CGA to include the galloyl conjugates of quinic acid and the cinnamoyl conjugates of quinic acid derivatives such as shikimic acid,^{23–26} methyl or butyl quinate,^{27,28} 4-deoxyquinic acid²⁹ and the recently characterised *muco*-quinic acid³⁰ which differs from the isomer described above by having an equatorial hydroxyl on carbon 3.

Other cinnamate conjugates

Cinnamic acids may be conjugated to many molecules other than quinic acid and its close relatives, although sometimes as very minor components of limited distribution. This second category includes:

- esters of other hydroxy acids, particularly αhydroxyhydrocaffeic, malic and tartaric but including galactaric, glucaric, gluconic, hydroxycitric, methoxyaldaric, phenylpyruvic and tartronic;
- (2) amides of amino compounds including aromatic amino acids, choline, anthranilic acids and diamines;
- (3) esters of polysaccharides, simple sugars, sugar alcohols including glycerol and myo-inositol, and glycosides including those of anthocyanins, flavonols and diterpenes;
- (4) esters of lipids including alkanols, alkandiols, ω -hydroxy-fatty acids and sterols; and
- (5) glycosides.

Cinnamate transformation products

There is little information on cinnamate transformation during food preparation and processing. It has been well established that cinnamic acids may be released from conjugates by hydrolysis and subseheat³¹ quently decarboxylated either by or microorganisms yielding various alkylphenols, but little consideration has been given to the possibility that food processing might generate novel transformation products. Possibly the first to be recognised was the grape reaction product (GRP) 2-S-glutathionylcaftaric acid and 2,5-di-S-glutathionylcaftaric acid formed from caftaric acid (caffeoyl tartaric acid) during wine making.^{32–35} Red wine contains adducts formed by the interaction of anthocyanins with vinyl phenol from p-coumarate decarboxylation^{36,37} and caffeic acid released during coffee roasting is partially converted to tetrahydroxy-phenylindans.38,39 These products will not be further discussed here as the cinnamate residues have been lost irretrievably.

Hydrolysis of CGA during coffee roasting also releases the 'typical' quinic acid residue (*vide supra*) which is then converted to the full theoretical complement of quinic acid and quinic lactone (quinide) diastereoisomers.^{40,41} Unhydrolysed CQA and FQA can be converted similarly to the corresponding 1,5lactones or quinides^{42–44} and it seems possible that other diastereoisomers, either of the lactones or the original acids, might also be formed.

CINNAMATES IN FOODS AND BEVERAGES

Quantitative data are presented on a fresh weight basis unless specified otherwise.

Coffee

Coffee beans are one of the richest dietary sources of CGA and for many consumers must be the major dietary source. The CGA in coffee and coffee products have been extensively reviewed^{31,45} and only a summary is presented here. Depending on the species, green coffee beans contain some 6–10% CGA on a dry matter basis. During roasting there is a progressive destruction and transformation of CGA with some 8–10% being lost for every 1% loss of dry matter but substantial amounts survive to be extracted into domestic brews and commercial soluble coffee powders.

It has been estimated⁴⁶ that a 200 ml cup of roast and ground coffee might supply from 20 mg CGA (weak brew, very dark roast) up to 675 mg CGA (strong brew, very pale roast robusta). More recent studies (Clifford and Balyaya, unpublished data) suggest that 70-200 mg per 200 ml cup of arabica and 70-300 (or even 350) mg per 200 ml cup of robusta are more representative. Soluble coffee powder as sold in the UK during 1983-1984 (2g per cup) might supply from 70 to 220 mg CGA consisting of 50-150 mg CQA, 4–21 mg FQA and 15–38 mg diCQA.^{31,47,48} Beverage prepared from Angolan robusta coffee beans, which have a unique composition, has been reported to contain some 12- $44\,\mathrm{mg}\,\mathrm{litre}^{-1}$ caffeoyl-tyrosine and some 33-66 mg litre⁻¹ of a similar but unidentified conjugate.49-51

The two major chlorogenic lactones (3-CQL and 4-CQL) are found at levels of 800-2400 and $200-1600 \text{ mg kg}^{-1}$ in roasted coffees.^{43,44} Coffee brew also contains traces of diterpene glycosides esterified with the relatively rare 3,4-dimethoxy-cinnamic acid.⁵²

Green and black tea

Tea leaf is the major dietary source of theogallin (5-GQA) and is accompanied by small amounts of pCoQA and CQA. These substances are not substrates for tea polyphenol oxidase and appear to survive fermentation and may thus be found in black tea as well as green tea. There are no data for their contents in tea brew, but 10–50 g kg⁻¹ has been reported for green and black tea leaf.^{53–56} Rooibos or bush tea (*Aspalathus linearis*), consumed in southern Africa, is quite different, but contains a novel conjugate between caffeic acid and the enol form of phenylpyruvic acid.⁵⁷

Maté

Maté is a traditional South American beverage prepared from the leaves of *Ilex paraguariensis* which is rich in CQA and diCQA. There have been few studies of the brew composition but green maté material bought and brewed in Europe provided 107–133 mg CGA per approx 200 ml of which diCQA represented approximately one third. In contrast, roasted maté provided 16–41 mg CGA per approx 200 ml.⁵⁸ Components originally assigned tentatively as FQA are now thought to be caffeoyl-quinic lactones (Clifford, unpublished data). Mazzafera⁵⁹ doubted the authenticity of the plant material purchased in Europe and reported that when authentic material was brewed in the traditional manner (50–60 g leaf per litre boiling water), some 480–520 mg 5-CQA and 340 mg caffeic acid were extracted. However, due to significant differences in the methods of analysis, these data are not directly comparable with the former.

Сосоа

The caffeoyl amide of dihydroxyphenylalanine and the *p*-coumaroyl amide of hydroxyphenylalanine have been reported in cocoa.⁶⁰

Pome fruits – apples and pears

Apples are one of the more extensively studied fruits. CGA are found at similar concentrations in the isolated flesh and skin (mean values $30-60 \text{ mg kg}^{-1}$) but are absent from the seeds and pomace.^{61,62} Whole apples have been reported to contain $62-385 \text{ mg kg}^{-1}$ CQA (with 5-CQA always dominant), up to 40 mg kg^{-1} pCoQA (with 4-pCoQA dominant in some cases) and smaller amounts of caffeoyl-, pcoumaroyl- and feruloyl-glucoses (up to 6, 6 and 9 mg kg^{-1} , respectively).⁶³⁻⁶⁵ Values of 100-200 mg kg⁻¹ total CGA on a dry matter basis are consistent with the foregoing.⁶⁶ It is clear that composition varies markedly with variety, cider varieties being richer than culinary. Commercial juices and ciders are also variable in composition with juices providing from undetectable up to 208 mg litre⁻¹ $CQA^{65,67}$ and ciders 11–480 mg litre⁻¹ (Clifford and Mitchell, unpublished data) or even higher when produced from single varieties of apples (for example, $>700 \text{ mg litre}^{-1} \text{ CQA and } >400 \text{ mg litre}^{-1}$ pCoQA for cv Vilberrie).⁶⁸ Heat processing reduced the CGA content of apple juice significantly, especially when accompanied by treatment with pectic enzymes, but the fate of the cinnamates was not discussed.⁶⁹ The fruit of Sorbus domestica, used in Germany as an adjunct in cider making, contains some $1500 \,\mathrm{mg \, kg^{-1}}$ CQA.⁷⁰

According to some investigators^{63-65,67} pears and pear juices are similar in composition with some 60– 280 mg kg⁻¹ CQA in whole fruit and up to 240 mg litre⁻¹ in juice but others⁷¹⁻⁷³ have also reported the presence of caffeoyl- and *p*-coumaroyl-L(-) malic acids (ie 2S malic acid) and 3,5-diCQA (1–8 mg kg⁻¹).

Stone fruits

Stone fruits contain CQA and pCoQA with mean contents in the range 150–600 mg kg⁻¹ and 3–

200 mg kg⁻¹, respectively, at least in cherries found mainly adjacent to the peel. Compared with apples and pears, the 3-isomers are more prominent relative to the 5-isomers. Cherries and plums are rich in pCoQA compared with peaches and apricots.⁶⁴ Studies on commercial juices and nectars (containing added sugar syrup)⁶⁵ are not entirely in agreement with the foregoing. Levels of 5-CQA were some 3-4× greater than 3-CQA in peach and apricot juice with total CQA reaching ≈9 and ≈4 mg litre⁻¹, respectively: pCoQA was not detected but some 0.5 mg litre⁻¹ of free cinnamic acids was noted. Heat treatment of peach juice caused significant losses of CQA and pCoQA especially when combined with pectic enzyme treatment.⁶⁹

Berry fruits

Berry fruits encompass several distinct botanical families and thus differ markedly in their composition. Relatively few quantitative data are available. Blueberries contain 0.5-2g kg⁻¹ CQA^{74,75} and only small amounts ($\approx 60 \,\mathrm{mg \, kg^{-1}}$ in total) of other conjugates, mainly glucosides. Blackcurrants supply some $140 \,\mathrm{mg \, kg^{-1}}$ of total conjugates in which COA dominate ($\approx 40\%$), pCoQA and caffeoyl-glucose account for some 15% each and feruloyl-glucose, pcoumaroyl-glucose and total cinnamate β -D-glucosides for some 8% each. Blackberries provide some 70 mg kg⁻¹ with CQA dominant. Raspberries, strawberries, redcurrants and gooseberries are poor sources by comparison with the berries mentioned above, providing mainly glucose esters and β -D-glucosides totalling some 20-30 mg kg⁻¹.^{74,76} Cranberries contain a complex series of sugar derivatives involving both ester and glycosidic linkages, sometimes both, and varying numbers of sugar residues.⁸ Bilberry wine has been reported to contain 50 mg litre⁻¹ of CGA.⁷⁵

Citrus fruits

Citrus fruits contain the comparatively rare conjugates of glucaric (feruloyl, *p*-coumaroyl and diferuloyl) and galactaric acids (feruloyl, *p*coumaroyl) and some associated lactones. These occur mainly in the peel (170–250, 27–62, 55– 67 mg kg⁻¹ in oranges, grapefruit and lemon respectively), and at ≈ 25 , ≈ 17 and $\approx 2 \text{ mg kg}^{-1}$ in the flesh.^{77,78} A range of other conjugates, involving sugars and anthocyanins, have also been reported.

The following data relate to the total cinnamates released by hydrolysis. The flavedo of oranges (and juice) supplied the following in bound form: ≈ 30 (8–18) mg kg⁻¹ sinapic, ≈ 130 (40–50) mg kg⁻¹ ferulic, ≈ 18 (8–12) mg kg⁻¹ *p*-coumaric and ≈ 4 (11–25) mg kg⁻¹ caffeic for which there is known to be a low recovery: the corresponding data for grapefruit were ≈ 50 (12–20), ≈ 152 (30–40), ≈ 72 (15–20) and ≈ 4 (5–8) mg kg⁻¹.^{79,80} Juice from blood oranges contains these same cinnamates at a total concentration in the range 50–160 mg litre⁻¹ again with ferulic

acid (30–64 mg litre⁻¹) dominant.⁸¹ Some workers have reported up to 2 mg litre⁻¹ 5-CQA and smaller amounts of free cinnamic acids and feruloylglucose in commercial orange juices and nectars (containing added sugar syrup).⁶⁷

Grapes and wines

Grapes are characterised by cis- and transcinnamovltartrates (ie esters of L(+)-tartaric acid, ie 2R3R tartaric acid), in the pulp and juice as well as the skin, and the classic chlorogenic acids are absent.82 As usual caffeoyltartaric (caftaric) acid dominates but is accompanied by p-coumaroyltartaric (coutaric), feruloyltartaric (fertaric) and trans-coutaric glucoside.83 The composition depends on the species and cultivar to a greater extent than season, ripeness and growing location, and may serve as a chemotaxonomic criterion^{84,85} but literature values vary considerably. Grape juice from red and white varieties of Vitis vinifera grown in America varied from ≈ 10 to $430 \,\mathrm{mg}\,\mathrm{litre}^{-1}$ caftaric in one study but up to 600 mg litre⁻¹ in another, with little variation between skin (150 mg kg^{-1}) and flesh (110 mg kg^{-1}) although the skin is usually richer: coutaric ranged to $55 \,\mathrm{mg}\,\mathrm{litre}^{-1}$ and fertaric to 16 mg litre⁻¹.^{65,67,85–87} Levels in grapes from Tuscany were in the range $2-7 \text{ mg kg}^{-1}$ caftaric, 0.4- 5 mg kg^{-1} coutaric and up to 3 mg kg^{-1} fertaric.⁸⁸ Juice from V aestivalis and V labrusca contained 1350 and $460 \,\mathrm{mg}\,\mathrm{litre}^{-1}$ caftaric respectively, whereas it was barely detectable in juice from Vrotundifolia.85

Levels of caftaric and coutaric decline markedly by oxidation during the wine-making process and become more variable but levels of fertaric change less. Wines are extremely variable, with contents increasing with time of skin contact.^{89,90} Some American wines contained 9–116, up to 9 and up to 16 mg litre⁻¹ for caftaric, coutaric and fertaric respectively. Some 43 mg litre⁻¹ caftaric was reported for some French wines,^{86,91} but only 2– 12 mg litre⁻¹ for some Spanish wines.^{92–94} Japanese Koshu wines contained appreciably higher concentrations of caftaric (40–304 mg litre⁻¹) and coutaric (28–127 mg litre⁻¹) although Japanese Semillon wines were very similar in this respect to French Semillon.⁹⁵

2-S-Glutathionylcaftaric acid has been reported in juices and wines at 6-8 mg litre⁻¹.^{65,93} Prolonged maturation can result in release of up to 3 or 4 mg litre^{-1} caffeic and *p*-coumaric acid by hydrolysis, and ethyl esters may form by transesterification.^{32-35,83,95} Red grapes also contain several 6-*O*-*p*-coumaroyl- and caffeoyl-glucosides of various anthocyanidins at levels which vary considerably with cultivar.^{88,96,97}

Other fruits

Pineapple is characterised by the presence of the caffeoyl- and *p*-coumaroyl-glycerols but no quanti-

tative data are available.⁵ Other workers have reported small amounts of free cinnamic acids, 5-*p*CoQA and feruloylglucose totalling about 3 mg litre⁻¹.⁶⁷ Some dates contain caffeoylshikimic acids.^{24,25} Kiwi fruit contains $\approx 11 \text{ mg kg}^{-1}$ conjugated caffeic acid, about a quarter of which is fibre-associated.⁹⁸

Brassica vegetables

3-CQA, 3-pCoQA and 3-FQA have been reported^{99,100} in the leafy Brassicas (kale, cabbage and Brussels sprouts) at levels of $6-120 \,\mathrm{mg \, kg^{-1}}$, up to 104 mg kg^{-1} and up to 37 mg kg^{-1} respectively along with smaller amounts of the 4-isomers and little or none of the 5-isomers. Significant amounts of sugar derivatives have also been reported with feruloyl glucose and sinapoyl glucose dominating and levels up to $350 \,\mathrm{mg \, kg^{-1}}$ in kale and up to $80 \,\mathrm{mg \, kg^{-1}}$ in the others and, in red cabbage and radish, cinnamoyl esters of anthocyanins. Broccoli contained about 60 mg kg^{-1} CGA plus about 20 mg kg⁻¹ sugar esters, but cauliflower and radish contained not more than 20 mg kg^{-1} of CGA and sugar esters combined. More recent studies¹⁰¹ indicate that some broccoli florets contain several mixed feruloylsinapoyl esters of gentiobiose totalling $> 300 \,\mathrm{mg \, kg^{-1}}$. Radish leaf contained a series of malic acid esters at levels totalling $240-550 \text{ mg kg}^{-1}$. The corresponding levels in tuber peel were 40- 120 mg kg^{-1} but only $2-23 \text{ mg kg}^{-1}$ in peeled tubers. Only traces of sugar and quinic acid esters were found.¹⁰² Sinapoyl-choline is a characteristic component of mustard seed (Sinapis alba and Brassica nigra = B juncea), mustard and mustard containing products but quantitative data are not available. Small amounts (up to 3 mg kg^{-1}) were found in some cabbages.99

Chenopodiaceae - spinach and beetroot

Spinach is dominated by p-coumaroyl-meso-tartaric acid at levels around 200 mg kg⁻¹ along with pcoumaroylmalic acid (25–30 mg kg⁻¹) and glucose esters totalling some 60–80 mg kg⁻¹. Beetroot root provides traces of quinic acid and glucose esters and the leaf slightly larger amounts of the glucose esters but the tartaric acid derivative found in spinach leaf was not detected.^{103,104} Cell wall-bound cinnamates are dealt with under fibre, vide infra.

Asteraceae

The Asteraceae is a very large group of plants which vary markedly in their profiles and contents of cinnamates. In some cases the profile does not differ with tissue for a given species (for example dandelion¹⁰⁵) but it should not be assumed that this is always the case.

The leaves of lettuce, endive and chicory provide quinic acid, malic acid and tartaric acid esters among which caffeic derivatives, especially dicaffeoyltartaric acid (chicoric acid), dominate. The total cinnamate contents were, respectively, $50-120 \text{ mg kg}^{-1}$, $200-500 \text{ mg kg}^{-1}$ and some 20 mg kg^{-1} .¹⁰⁴ CQA and

diCQA have been found in chicory root (total $2.6 \,\mathrm{g \, kg^{-1}}$) and in dandelion and chicory root-based coffee substitutes (some 100–400 mg kg⁻¹).^{106,107} Although these workers saw no evidence of mono- or di-caffeoyltartaric acid in the coffee substitutes these have been found throughout all parts of dandel-ion.¹⁰⁵ Another study found caffeoylshikimic acid but not the quinic acid esters referred to above in endive.²⁶

Artichoke (Cynara scolymus) and scorzonera provide some 450 and 180 mg kg^{-1} CQA respectively with little or none of the other conjugates.¹⁰⁰ Artichoke is characterised by significant amounts of 3.5diCQA and 1,5-diCQA.¹⁰⁸⁻¹¹⁰ These isomers are not easily resolved and 1,5-diCQA is unstable in aqueous solutions, isomerising rapidly to 1,3-diCQA, otherwise known as cynarin(e). These factors may account for Herrmann⁸⁶ failing to mention these isomers although cynarin forms 800-1300 mg kg⁻¹ dry basis of the capitula.¹¹¹ It is not known whether any cinnamates occur in the artichoke-based beverage marketed as Cynar, but cynarin is found in some herbal medicines.¹¹² The relatively scarce triCQA, tetraCQA and caffeoylsuccinoyl-quinic acids (vide supra) have been reported in Asteraceae used as herbal medicines. These conjugates have not been reported in food plants other than burdock (Arctium lappa),^{20,21} an extract of which was once used in preparing the cordial 'Dandelion and Burdock'. Absinthe (Artemesia absinthum), tarragon (A dracunculus) and mugwort (A vulgaris) contain significant amounts of imperfectly defined quinic acid esters of caffeic acid¹¹³ but it is not known if these are transferred to vermouth and similar beverages in which extracts of absinthe are traditionally used.

Solanaceae

Potato tubers provide some $500-1200 \text{ mg kg}^{-1} \text{ dry}$ basis total CGA^{114,115} (mainly COA plus a little diCQA¹¹⁶) at harvest and the content rises slowly on storage especially in the light¹¹⁷ and during wound healing.¹¹⁸ These are concentrated in the peel where they may reach levels of $2-5 \text{ g kg}^{-1}$ (Ref 119) or even higher.¹²⁰ p-Coumaroyl conjugates of anthocyanins are present in peel (up to $7 g k g^{-1}$) and flesh (up to 2 g kg⁻¹).¹¹⁹ Bell peppers contain caffeoyl and feruloylputrescine in the placenta during fruit development but only small amounts of glucose derivatives in mature fruit. Aubergines have a significant CQA content (600 mg kg $^{-1}$) and small amounts of FQA and anthocyanin conjugates (15 mg kg^{-1}) . During the ripening of tomatoes, CQA in the pulp are partially replaced by glucosides in the peel - ripe fruit contain some 10-80 mg kg⁻¹ CQA, 5-15, 15-50 and $20-70 \,\mathrm{mg \, kg^{-1}}$ glucosides of ferulic, caffeic and pcoumaric respectively, plus up to $20 \,\mathrm{mg \, kg^{-1}}$ pcoumaroyl-glucose.^{104,116,121} Caffeoylglucaric acid is found in tomato leaf.¹²² It has not been possible to trace any compositional data for tomato juice or purée.

Apiaceae

Celery contains a modest but variable amount of CQA (2–65 mg kg⁻¹), pCoQA (up to 3 mg kg⁻¹) and FQA (up to 8 mg kg⁻¹),^{123,124} as do carrots (20–120 mg kg⁻¹)¹⁰⁰ but it should be noted that seedsmen attribute the root fly-resistance of modern varieties to the virtual absence of CGA.

This botanical group also supplies a range of spices and some of these have substantial contents of quinic acid conjugates (predominantly CQA but with appreciable FQA and pCoQA) and reaching $3 g k g^{-1}$ in the case of anise, just over $2 g k g^{-1}$ in fennel, approaching $2 g k g^{-1}$ in caraway, but only some 200 mg kg^{-1} in coriander.¹²⁵

Lamiaceae and Boraginaceae

The Lamiaceae supply many leafy herbs (basil, marjoram, melissa, oregano, peppermint, rosemary, sage, spearmint and thyme) characterised by the occurrence of rosmarinic acid, the caffeic acid conjugate of α -hydroxyhydrocaffeic acid, at concentrations ranging from 1 to 17 g kg^{-1} dry basis¹¹³ or even in excess of 20 g kg^{-1} (dry basis) in some samples of basil and rosemary.^{126,127} A smaller quantity (500 mg kg⁻¹ dry basis) has been found in borage.¹¹³ Melissa also contains a series of dimeric and trimeric caffeic acid conjugates.¹²⁸ It has not been possible to trace data for mixed herbs, stuffings, forcemeats, etc.

Fabaceae – legumes and pulses

There have been various reports of cinnamates in legumes and pulses $2^{3-26,129}$ but the distribution is variable across tissues, at least in Faba (Phaseolus spp) type beans where the largest concentrations of L(-)-malic acid conjugates (phaseolic acids) are found in leaves, with moderate but variable contents (up to 100 mg kg^{-1}) in pods and traces only in the seeds.¹⁰⁴ Peas contain p-coumaroyl, caffeoyl and feruloyl derivatives of flavonol glycosides¹³⁰ and lentils 0.1 mg kg⁻¹ of bound *p*-coumaric acid.⁹⁸ Caffeoyl and p-coumaroyltartronic acids have been found dry mung in beans (Vigna radicata = Phaseolus aureus).¹³¹

The novel compound, 1-caffeoyl-4-deoxyquinic acid, along with 3-CQA and 5-CQA, has been reported in the foliage of a wild groundnut (*Arachis paraguariensis*) but it is not clear if these compounds occur also in the edible portions of the *A hypogea*.²⁹ Commercial peanut flour and protein isolates have significant cinnamate contents including 500–1350 mg kg⁻¹ *p*-coumaric acid.⁶⁹

Miscellaneous vegetables

Corn salad (Valerianaceae) contains approximately 1 g kg^{-1} CQA. Rhubarb petiole (*Rheum* spp, Polygonaceae) contains traces of glucose esters but the leaf contains *p*CoQA as the only CGA. Leeks (Alliaceae) contain small amounts of FQA (up to 12 mg kg^{-1}) without detectable CQA.¹⁰⁰ The spice star anise contains from 100 to 200 mg kg⁻¹ CGA in which, surprisingly, 4-*p*CoQA dominates at 52–

125 mg kg⁻¹.¹²⁵ Sweet potatoes (*Ipomoea batatas*) contain CQA and diCQA in the range 100– 500 mg kg⁻¹,¹³² a series of long chain (C_{16} , C_{17} , C_{18}) alkyl ferulates¹³³ and in pigmented varieties caffeoyl derivatives of anthocyanins.¹³⁴ Redpigmented yams (*Dioscorea* spp) contain sinapic esters of a cyanidin glycoside.¹³⁵ Chinese water chestnuts (*Eleocharis dulcis*, Cyperaceae) contain cell wall-bound cinnamates, *vide infra*.

Cereals and cereal products including beers and bran

It has not been possible to trace any data for bread, biscuits, cakes, breakfast cereals, etc.

Conjugates of cell wall polysaccharides

Cereals are characterised by cinnamic acids, especially ferulic acid, esterified to arabinose residues in primary cell wall arabinoxylan in the aleurone layer and pericarp. A portion of this total cinnamate exists as ferulate dimers linked in various ways including 5,5' or 5,8' carbon-carbon bonds.¹³⁶⁻¹³⁸

Barley bran contains $\approx 50 \text{ mg kg}^{-1}$ bound ferulic, $\approx 30 \text{ mg kg}^{-1}$ bound *p*-coumaric and 3 mg kg^{-1} diferulic acid. The endosperm content is $\approx 3 \text{ mg kg}^{-1}$ total and the aleurone is intermediate.⁶⁹ Rice endosperm cell walls contain 12 g kg^{-1} esterified cinnamic acids comprising $\approx 9 \text{ g kg}^{-1}$ ferulic, $\approx 2.5 \text{ g kg}^{-1}$ *p*-coumaric and $\approx 0.5 \text{ g kg}^{-1}$ diferulic esters.¹³⁹ While whole wheat contains some 20– 30 mg kg^{-1} cinnamic acids esterified to polysaccharides,¹⁴⁰ the derived wheat bran contains some $4-7 \text{ g kg}^{-1}$ and maize bran as much as $30 \text{ g kg}^{-1.140-142}$ Maize leaves contain conjugates of hydroxycitric acid.¹⁴³

Water chestnuts (*Eleocharis dulcis*, Cyperaceae) are botanically closer to the cereals than to other common fruits and vegetables and are characterised also by a significant content of cell wall-bound cinnamates ($>7 \text{ g kg}^{-1}$ ferulate and $>4.5 \text{ g kg}^{-1}$ diferulate, the majority of which is 8-O-4 linked).¹⁴⁴

Spinach and sugar beet contain cell wall polysaccharides which are esterified with *p*-coumaric and ferulic acids, and to a lesser extent with what is probably cross-linked ferulate dimer.^{142,145} Sugar beet fibre contains some $9 g k g^{-1}$ of ferulic acid attached to galactose or arabinose residues in pectic sidechains. Digestion, using an *in vitro* model of the human colon suggests that some 90% of cinnamates are released from sugar beet fibre in 24h compared with some 35–40% from wheat bran but the released cinnamate may be further metabolised by the gut flora and its bioavailability is uncertain.¹⁴²

Other conjugates and free cinnamates

Wheat, maize and rye contain ferulic and *p*-coumaric esters of sterols and stanols.^{146,147} The leaves of rye contain feruloyl esters of gluconic and methoxy-aldaric acids.¹⁴⁸ Oats contain a series of 24 caffeic and ferulic esters of glycerol, long chain alkanols, alkandiols (n = 22, 23, 24) and ω -hydroxy acids

(n = 26, 28).^{149–153} In addition, there are more than 25 compounds which are esters of anthranilic acid or 5-hydroxyanthranilic acid with either *p*-coumaric, caffeic or ferulic acids (avenanthramides) or with their ethylenic analogues (avenulamides).^{154–156} Oat meal has been reported to contain some 200–300 mg kg⁻¹ of esterified ferulic acid.¹⁴⁰

Barley contains small amounts of free cinnamic acids, mainly *p*-coumaric and ferulic which together range from 8 to 26 mg kg^{-1} (Ref 157) but this value rises during malting (18 mg kg^{-1}). The content rises further on mashing, presumably by hydrolysis of bound forms, with ferulic acid dominating (115 mg kg^{-1}) and lesser but equal amounts of caffeic and *p*-coumaric making up the total (167 mg kg^{-1}). A limited range of beers (including lagers, stouts, ales, etc) contained 2–8 mg litre⁻¹ but German 'rauchbier' reached almost 20 mg litre⁻¹.¹⁵⁷

DIETARY BURDEN

A major objective of this review, as originally conceived, was to identify the commodities most likely to provide the greatest amount of (i) a particular cinnamate (independent of the conjugate type and extent of hydrolysis in the gastro-intestinal tract) and (ii) of a particular conjugate. These two approaches have been taken since it seems possible that the form of the conjugate might influence absorption as is suggested in the case of flavonol glycosides.¹⁵⁸ With the exception of some beverages, the lack of data for the composition of commodities as consumed, ie the edible portion after processing, cooking, baking, etc, effectively make such an assessment impossible. In the circumstances, it is possible only to highlight the sources which have the potential in normal usage to make a greater contribution than most.

The values for burden set out below have been calculated from representative but rounded quantitative data presented in the first part of this review and assuming, for simplicity, that a serving of fruit or vegetable is 100 g, of non-alcoholic beverage is 200 ml, of cider or beer is 500 ml, of wine is 100 ml, of bran is 10 g and of herbs is 1 g.

There is no doubt that caffeic acid is the cinnamate which occurs most extensively, and the various CQA and diCQA are the most ubiquitous conjugates. Usually the 5-isomer dominates, but in stone fruit and brassicas it is the 3-isomer. Because of the quantity commonly consumed, coffee beverage must top the list with 200 ml instant brew (2% w/v) supplying 50–150 mg CQA. The theoretical yield of caffeic acid would be $\approx 25-75$ mg. Blueberries, aubergines, apples, cider and green maté could be important in some populations. Sweet cherries and Savoy cabbage along with apples, apple juice and cider probably supply the majority of the *p*CoQA while coffee must, in the majority of cases, come first for conjugated ferulic acid, followed by citrus juices.

In view of the quantities consumed by some populations, wines may be the major source of the tartaric acid conjugates, but the literature data are extremely variable probably reflecting differences in winemaking practice across the world. Clearly grapes and grape juice also could make a significant contribution especially of caftaric at up to 60 mg per 200 ml juice. Lettuce, with 5–15 mg $100 g^{-1}$, will be the major source of chicoric (dicaffeoyltartaric) acid in the UK, but endive may have twice the concentration. Spinach will be the major source of coutaric (albeit *meso*-tartaric) and is almost certainly the richest source of conjugated *p*-coumaric acid at some 30– 35 mg per 100 g.

Lettuce, since it is eaten raw, will probably be the main UK source of caffeoylmalic acid (up to $3 \text{ mg} 100 \text{ g}^{-1}$) but spinach may be richer. Legumes might also be an important source but the quantitative data are inadequate.

Broccoli florets and leafy cruciferous vegetables will be the major source of sugar esters and of conjugated sinapic acid – broccoli florets supplying >10 mg bound sinapic acid per 100 g. Tomatoes and tomato products are likely to be the major source of glucosides at up to $13 \text{ mg } 100 \text{ g}^{-1}$ in total, and possibly the second richest source of conjugated *p*coumaric acid at $\approx 3 \text{ mg } 100 \text{ g}^{-1}$.

Herbs are the only source of rosmarinic acid. While some are very rich, the levels consumed make this a comparatively poor source of conjugated caffeic acid, generally supplying only low milligram quantities per gram, but up to $\approx 10 \text{ mg}$ in some cases.

Cereal bran and bran-enriched products must be the most important source of cell wall-bound cinnamates. Even with small servings, say 10 g, maize or wheat bran could supply up to 30 and 7 mg of ferulate respectively making such products the richest dietary source of this cinnamate although its bioavailability after release in the colon is unclear. However, coffee brew could supply up to 10 mg ferulate (as FQA) per 200 ml cup so, for many people, coffee will be the major source.

There are insufficient quantitative data to allow useful comments to be made about the other conjugates.

CONCLUDING REMARKS

Cinnamate content of foods and beverages is incompletely known yet variable, as are diets. For a meal consisting of two glasses of wine, salad-based starter, main course with potatoes and two other vegetables including broccoli or spinach, a blueberry sweet, plus two large cups of strong coffee, the intake might be as much as 1g of conjugates (predominantly CGA) or 500 mg of cinnamates (predominantly caffeic). The coffee alone would supply approximately 70% of the total *even if there were no losses in cooking*. In contrast, breakfast consisting of two slices of white toast and a cup (or two) of tea would supply hardly any cinnamates, but replacing white bread with wholemeal, taking a bran-containing cereal and a glass of citrus juice might supply up to some 60 mg *cinnamates* (there are inadequate data for conjugates contents) in which ferulate would dominate.

It seems likely that the UK population will fall into several categories depending on (i) their consumption of coffee, (ii) their consumption of bran, and (iii) their consumption of citrus. Those who drink several cups of coffee per day augmented by bran and citrus might easily achieve 500–800 mg *cinnamates* (or even 1g for the highest coffee consumption) whereas those who eschew all these and take little fresh fruit or vegetables might struggle to achieve 25 mg.

ACKNOWLEDGEMENTS

The author gratefully acknowledges the advice of Dr Véronique Cheynier (INRA Institut des Produits de la Vigne, Montpellier, France) and Dr Tino Santos-Buelga (Departmento de Química Analítica, Universidad de Salamanca, Spain) in preparing the section on grapes and wines.

REFERENCES

- 1 Robiquet and Boutron, Ueber den Kaffee von Robiquet und Boutron. Ann Pharmacie 23:93–95 (1837).
- 2 Payen, Untersuchung des Kaffees. Annalen 60:286–294 (1846).
- 3 Payen, Memoire sur le café (3° Part). Comptes Rendus 23:244–251 (1846).
- 4 Fischer HOL and Dangschat G, Konstitution der Chlorogensäure (3. Mitteil. über Chinasäure und Derivate). Berichte 65:1037–1040 (1932).
- 5 Takata RH and Scheuer PJ, Isolation of glyceryl esters of caffeic and *p*-coumaric acids from pineapple stems. *Lloydia* **39**:409–411 (1976).
- 6 Baranowski JD and Nagel CW, Isolation and identification of the hydroxycinnamic acid derivatives in white Riesling wine. Am J Enol Vitic 32:5-13 (1981).
- 7 Clifford MN, Coffee bean dicaffeoylquinic acids. Phytochemistry 25:1767-1769 (1986).
- 8 Marwan AG and Nagel CW, Separation and purification of hydroxycinnamic acid derivatives in cranberries. *J Food Sci* 47:774–778, 782 (1982).
- 9 IUPAC, Nomenclature of cyclitols. *Biochem J* **153**:23-31 (1976).
- 10 Timmerman BN, Hoffmann JJ, Jolad SD, Schram KH, Klenk RE and Bates RB, Constituents of *Chrysothamnus* paniculatus. 3. 3,4,5-tricaffeoylquinic acid (a new shikimate prearomatic) and 3,4-, 3,5- and 4,5-dicaffeoylquinic acids. *J Nat Prod* 46:365-368 (1983).
- 11 Merfort I, Caffeoylquinic acids from flowers of Arnica montana and Arnica chamossonis. Phytochemistry 31:2111-2113 (1992).
- 12 Scholz E, Heinrich M and Hunkler D, Caffeoylquinic acids and some biological activities of *Pluchea symphytifolia*. *Planta Med* **60**:360–364 (1994).
- 13 Peluso G, de Feo V, de Simone F, Bresciano E and Vuotto ML, Studies on the inhibitory effects of caffeoylquinic acids on monocyte migration and superoxide ion production. *J Nat Prod* 58:639-646 (1995).
- 14 Agata I, Goto S, Hatano T, Ishibe S and Okuda T, 1,3,5-Tri-O-caffeoylquinic acid from Xanthium strumarium Phytochemistry **33**:508-509 (1997).
- 15 Iwahashi H, Morishita H, Osaka N and Kido R, 3-Oferuloyl-4-O-caffeoylquinic acid from coffee beans. Phytochemistry 24:630-632 (1985).
- 16 Morishita H, Iwahashi H and Kido R, 3-O-Caffeoyl-4-O-

feruloylquinic acid from green robusta coffee beans. *Phyto-chemistry* 25:2679–2680 (1986).

- 17 Clifford MN, Kellard B and Birch GG, Characterisation of caffeoylferuloylquinic acids by simultaneous isomerisation and transesterification with tetramethylammonium hydroxide. *Food Chem* 34:81–88 (1989).
- 18 Balyaya KJ and Clifford MN, Individual chlorogenic acids and caffeine contents in commercial grades of wet and dry processed Indian green robusta coffee beans. *J Food Sci Technol Mysore* 32:104–108 (1995).
- 19 Nishizawa M, Izuhara R, Kaneko K and Fujimoto Y, 3-Caffeoyl-4-sinapoylquinic acid, a novel lipoxygenase inhibitor from *Gardeniae fructus*. Chem Pharm Bull Tokyo 35:2133 (1987).
- 20 Tsuchida T, Suzuki M and Kurogi M, Evaluation of antioxidant activity of vegetable extracts and determination of some active compounds. *Nippon Kogyo Shokuhin Gakkaishi* 13:199–206 (1968).
- 21 Maruta Y, Kawabata J and Niki R, Antioxidative caffeoylquinic acid derivatives in the roots of Burdock (Arctium lappa L.). J Agric Food Chem 43:2592-2595 (1995).
- 22 Chuda Y, Ono H, Ohnishi-Kameyama M, Nagata T and Tsushida T, Structural identification of two antioxidant quinic acid derivatives from Garland (Chrysanthemum coronarium L). J Agric Food Chem 44:2037-2039 (1996).
- 23 Fukuoka M, Chemical and toxicological studies on Bracken Fern (*Pteridium aquilinum*) var. *latiusculum*. VI. Isolation of 5-O-caffeoylshikimic acid as an antihistamine factor. *Chem Pharm Bull Tokyo* 10:3219–3224 (1962).
- 24 Maier VP, Metzler DM and Huber AF, 3-O-caffeoylshikimic acid (dactylifric acid) and its isomers, a new class of enzymic browning substrates. *Biochem Biophys Res Commun* 14:124–128 (1964).
- 25 Harborne JB, Williams CA and Greenham J, Distribution of charged flavones and caffeoylshikimic acid in Palmae. *Phytochemistry* 13:1557–1559 (1974).
- 26 Goupy PM, Varoquaux PJA, Nicolas JJ and Macheix JJ, Identification and localization of hydroxycinnamoyl and flavonol derivatives from endive (*Cichorium endivia* L cv Geante Maraichere) leaves. J Agric Food Chem 38:2116– 2121 (1990).
- 27 Barron D, Kaouadji M and Mariotte A-M, New cinnamoyl esters of quinic acid from *Meum athamanticum*. Z Naturforsch **39:1**67–170 (1984).
- 28 Rumbero-Sanchez A and Vazquez P, Quinic acid esters from Isertia haenkeana. Phytochemistry 30:311-313 (1991).
- 29 Stevenson PC, Anderson JC, Blaney M and Simmonds MSJ, Developmental inhibition of Spodoptera litura (Fab) larvae by a novel caffeoylquinic acid from wild groundnut Arachis paraguariensis (Chod et Hassl). J Chem Ecol 19:2917–2933 (1993).
- 30 Haribal M, Feeny P and Lester CC, Caffeoylcyclohexane-1carboxylic acid derivative from Asimina triloba. Phytochemistry 49:103-108 (1998).
- 31 Clifford MN, Chlorogenic acids, in Coffee 1. Chemistry. 1 Ed by Clarke RJ and Macrae R. Elsevier Applied Science, London, UK, pp 153–202 (1985).
- 32 Singleton VL, Zaya J, Trousdale E and Salgues M, Caftaric acid in grapes and conversion to a reaction product during processing. *Vitis* 23:113–120 (1984).
- 33 Singleton VL, Salgues M, Zaya J and Trousdale E, Caftaric acid disappearance and conversion to products of enzymic oxidation in grape must and wine. Am J Enol Vitic 36:50– 56 (1985).
- 34 Cheynier V, Trousdale EK, Singleton VL, Salgues MJ and Wylde R, Characterization of 2-S-glutathionylcaftaric acid and its hydrolysis in relation to grape wines. J Agric Food Chem 34:217-221 (1986).
- 35 Salgues M, Cheynier V, Gunata Z and Wylde R, Oxidation of grape juice 2-S-glutathionyl caffeoyl tartaric acid by *Botrytis cinerea* laccase and characterization of a new substance: 2,5-di-S-glutathionyl caffeoyl tartaric acid. *J Food*

Sci 51:1191-1194 (1986).

- 36 Fulcrand H, Cameira-dos-Santos P-J, Sarni-Manchado P, Cheynier V and Favre-Bonvin J, Structure of new anthocyanin-derived wine pigments. J Chem Soc Perkin Trans 1 735-739 (1996).
- 37 Fulcrand H, Cameira-dos-Santos P-J, Sarni-Manchado P, Cheynier V and Moutounet M, New anthocyanin-derived wine pigments. in *Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols.* 2, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF. INRA Editions, Paris, France, pp 259–260 (1996).
- 38 Guillot FL, Malnoe A and Stadler RH, Antioxidant properties of novel tetraoxygenated phenylindan isomers formed during thermal decomposition of caffeic acid. J Agric Food Chem 44:2503-2510 (1996).
- 39 Stadler RH, Welti DH, Stämpfli AA and Fay LB, Thermal decomposition of caffeic acid in model systems. Identification of novel tetraoxygenated phenylindan isomers and their stability in aqueous solution. *J Agric Food Chem* 44:898–905 (1996).
- 40 Scholz BM and Maier HG, Isomers of quinic acid and quinide in roasted coffee. Z Lebensm Unters Forsch 190:132-134 (1990).
- 41 Scholz-Böttcher BM, Ludger E and Maier HG, New stereoisomers of quinic acid and their lactones. *Liebigs Ann Chim* 1029–1036 (1991).
- 42 Scholz-Böttcher BM, Bildung von Säuren und Lactonen insbesondere aus Chlorogensäure, beim Rösten von Kaffee. Braunschweig, Germany (1991).
- 43 Bennat C, Engelhardt UH, Kiehne A and Wirries FM, HPLC analysis of chlorogenic acid lactones in roasted coffee. Z Lebensm Unters Forsch 199:17-21 (1994).
- 44 Schrader K, Kiehne A, Engelhardt UH and Maier HG, Determination of chlorogenic acids with lactones in roasted coffee. J Sci Food Agric 71:392–398 (1996).
- 45 Clifford MN, The nature of chlorogenic acids are they advantageous compounds in coffee? in *Dix-septième Colloque Scientifique Internationale sur le Café*, ASIC, Paris, France (1998).
- 46 Clifford MN and Walker R, Letter to the editor: Chlorogenic acids – confounders of coffee-serum cholesterol relationships. *Food Chem* 24:77–80 (1987).
- 47 Trugo LC and Macrae R, Chlorogenic acid composition of instant coffees. Analyst 109:263–266 (1984).
- 48 Clifford MN and Willson KC, Coffee: Botany, Biochemistry and Production of Beans and Beverage, Chapman and Hall, London, UK, p 457 (1985).
- 49 Clifford MN, Kellard B and Ah-Sing E, Caffeoyl-tyrosine from green robusta coffee beans. *Phytochemistry* 28:1989– 1990 (1989).
- 50 Correia AMNG, Leitao MCA, Phenolic acids in green and roasted coffee, in XVIe Journées Internationales Groupe Polyphénols and The Royal Society of Chemistry, Lisbon, 2, DTA, Lisbon, Portugal, pp 117-120 (1992).
- 51 Correia AMNG, Leitao MCA and Clifford MN, Caffeoyltyrosine and Angola II as characteristic markers for Angolan robusta coffees. *Food Chem* 53:309–313 (1995).
- 52 Maier HG and Wewetzer H, Bestimmung von Diterpen-Glykosiden im Bohnenkaffee. Z Lebensm Unters Forsch 167:105-107 (1978).
- 53 Roberts EAH and Myers M, Theogallin, a polyphenol occurring in tea. II. Identification as a galloylquinic acid. J Sci Food Agric 9:701-705 (1958).
- 54 Hara Y, Luo S-J, Wickremasinghe RL and Yamanishi T, Chemical composition of tea. *Food Rev Int* 11:435–456 (1995).
- 55 Shao W, Powell C and Clifford MN, The analysis by HPLC of green, black and Pu'er teas produced in Yunnan. J Sci Food Agric 69:535-540 (1995).
- 56 Kiehne A and Engelhardt UH, Thermospray-LC-MS analysis of various groups of polyphenols in tea. II: Chlorogenic acids, theaflavins and thearubigins. Z Lebensm

Unters Forsch 202:299-302 (1996).

- 57 Marais C, Ferreira D and Steenkamp JA, Phenolic metabolites from Rooibos Tea (Aspalathus linearis), in Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols 2, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF, INRA Editions, Paris, France, pp 333–334 (1996).
- 58 Clifford MN and Ramirez-Martinez JR, Chlorogenic acids and purine alkaloid content of Maté (*Ilex paraguariensis*) leaf and beverage. *Food Chem* 35:13–21 (1990).
- 59 Mazzafera P, Maté drinking: caffeine and phenolic acid intake. Food Chem 60:67-71 (1997).
- 60 Sanbongi C, Osakabe N, Natsume M, Takizawa T, Gomi S and Osawa T, Antioxidative polyphenols isolated from *Theobroma cacao. J Agric Food Chem* **46**:454–457 (1998).
- 61 Lu Y and Foo Y, Identification and quantification of major polyphenols in apple pomace. *Food Chem* **59**:187–194 (1997).
- 62 Burda S, Oleszek W and Lee CY, Phenolic compounds and their changes in apples during maturation and cold storage. *J Agric Food Chem* 38:945-948 (1998).
- 63 Mosel HD and Herrmann K, The contents of catechins and hydroxycinnamic acids in pome and stone fruits. Z Lebensm Unters Forsch 154:6–11 (1974).
- 64 Risch B and Herrmann K, Die Gehalte an Hydroxyzimtsäure-Verbindungen und Catechinen in Kernund Steinobst. Z Lebensm Unters Forsch 186:225–230 (1988).
- 65 Spanos GA and Wrolstad RE, Phenolics of apple, pear, and white grape juices and their changes with processing and storage – a review. J Agric Food Chem 40:1478–1487 (1992).
- 66 Amiot MJ, Tacchini M, Aubert S and Nicolas J, Phenolic composition and browning susceptibility of various apple cultivars at maturity. *J Food Sci* 57:958–962 (1992).
- 67 Fernández de Simon B, Pérez-Ilzarbe J, Hernández T, Gómez-Cordovés C and Estrella I, Importance of phenolic compounds for the characterization of fruit juices. J Agric Food Chem 40:1531-1535 (1992).
- 68 Whiting GC and Coggins RA, Estimation of the monomeric phenolics of ciders. J Sci Food Agric 26:1833–1838 (1975).
- 69 Hernández T, Ausin N, Estrella I, Gomez-Cordoves C, Bartolomé B, Bengoechea L and Perez-Ilzarbe FJ, Phenolic compounds in fruit juices. Variations with different treatments, in *Polyphenols 94. XVIIe Journeés Internationales Groupe Polyphénols*, Ed by Brouillard R, Jay M and Scalbert A. INRA Editions, Paris, France, pp 221-222 (1995).
- 70 Ritter G, Hagenauer-Hener U and Dietrich H, The phenolic substances of Sorbus domestica L and their importance for the quality of apple wines, in Polyphenols 94. XVIIe Journeés Internationales Groupe Polyphénols, Ed by Brouillard R, Jay M and Scalbert A. INRA Editions, Paris, France, pp 235-236 (1995).
- 71 Wald B, Wray V, Galensa R and Herrmann K, Malonated flavonol glycosides and 3,5-dicaffeoylquinic acid from pears. *Phytochemistry* **28**:663-664 (1989).
- 72 Oleszek W, Amiot MJ and Aubert S, Identification of some phenolics in pear fruit. J Agric Food Chem 42:1261–1265 (1994).
- 73 Kodoma M, Wada H, Otani H, Kohmoto K and Kimura Y, 3,5-di-O-caffeoylquinic acid, an infection-inhibiting factor from Pyrus pyrifolia induced by infection with Alternaria alternata. Phytochemistry 47:371-373 (1997).
- 74 Schuster B and Herrmann K, Hydroxybenzoic and hydroxycinnamic acid derivatives in soft fruits. *Phytochemistry* 24:2761-2764 (1985).
- 75 Gao L and Mazza G, Quantitation and distribution of simple and acylated anthocyanins and other phenolics in blueberries. *J Food Sci* 59:1057–1059 (1994).
- 76 Koeppen BH and Herrmann K, Phenolics of fruits. IX. Flavonoid glycosides and hydroxycinnamic acid esters of blackcurrants (*Ribes nigrum*). Z Lebensm Unters Forsch 164:263-268 (1977).

- 78 Risch B and Herrmann K, Contents of hydroxycinnamic acid derivatives in citrus fruits. Z Lebensm Unters Forsch 187:530-534 (1988).
- 79 Peleg H, Naim M, Rouseff RL and Zehavi U, Distribution of bound and free phenolic acids in oranges (*Citrus sinensis*) and grapefruits (*Citrus paradisi*). J Sci Food Agric 57:417– 426 (1991).
- 80 Naim M, Zehavi U, Nagy S and Ronseff RL, Hydroxycinnamic acids as off-flavour precursors in citrus fruits and their products, in *Phenolic Compounds in Foods and their Effects on Health. 1 Analysis, Occurrence and Chemistry. No.* 506, Ed by Ho C-T, Lee CY and Huang M-T. American Chemical Society, Washington, DC, USA, pp 180–191 (1992).
- 81 Rapisarda P, Carollo G, Fallico B, Tomaselli F and Maccarone E, Hydroxycinnamic acids as markers of Italian blood orange juices. *J Agric Food Chem* 46:464–470 (1998).
- 82 Singleton VL, Timberlake CF and Lea AGH, The phenolic cinnamates of white grapes and wine. J Sci Food Agric 29:403-310 (1978).
- 83 Somers TC, Verette E and Pocock KF, Hydroxycinnamate ester of Vitis vinifera changes during vinifications, and effect of exogenous enzymic hydrolysis. J Sci Food Agric 40:67-78 (1987).
- 84 Boursiquot JM, Sapis JM and Macheix JJ, Les esters hydroxycinnamiques chez le genre Vitis. Essais d'application taxonomique premiers résultats. CR Acad Sci Paris Series III 302:177 (1986).
- 85 Singleton VL, Zaya J and Trousdale EK, Caftaric and coutaric acids in fruit of *Vitis*. *Phytochem* 25:2127–2133 (1986).
- 86 Herrmann K, Occurrence and content of hydroxycinnamic and hydroxybenzoic acid compounds in foods. CRC Crit Rev Food Sci Nutr 28:315–347 (1989).
- 87 Lee CY and Jaworski A, Major phenolic compounds in ripening white grapes. Am J Enol Vitic 40:43-46 (1989).
- 88 Baldi A, Romani A, Mulinacci N and Vincieri FF, Composés phenoliques dans les cépages de Toscane de Vitis vinifera L. J Int Sci Vigne Vin 27:201–215 (1993).
- 89 Cheynier V, Rigaud J, Souquet J-M, Barillere JM and Moutounet M, Effect of pomace contact and hyperoxidation on the phenolic composition and quality of Grenache and Chardonnay wines. Am J Enol Vitic 40:36–42 (1989).
- 90 Ricardo-da-Silva JM, Cheynier V, Samson A and Bourzeix M, Effect of pomace contact, carbonic maceration, and hyperoxidation on the procyanidin composition of Grenache blanc wines. Am J Enol Vitic 44:168–172 (1993).
- 91 Herrick IW and Nagel CW, The caffeoyltartrate content of White Riesling wines from California, Washington, and Alsace. Am J Enol Vitic 36:95–97 (1985).
- 92 Andres-Lacueva C, Lamuela-Raventós RM, Buxaderas S and Torre-Boronat MC, Polyphenol content in spakling wines (Cava) at different aging period, in *Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols.* 2, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF. INRA Editions, Paris, France, pp 305-306 (1996).
- 93 Bétes-Saura C, Andrés-Lacuva C and Lamuela-Raventós RM, Phenolics in white free run juices and wines from Penedès by high-performance liquid chromatography: changes during vinification. J Agric Food Chem 44:3040– 3046 (1996).
- 94 Lao C, López-Tamames E, Lamuela-Raventós RM, Buxaderas S and Torre-Boronat MC, Effect of grape pectic enzyme treatment on phenolics of white musts and wines, in *Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols.* 2, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF. INRA Editions, Paris, France, pp 303–304 (1996).
- 95 Okamura S and Watanabe M, Determination of phenolic cinnamates in white wine and their effect on wine quality.

Agric Biol Chem 45:2063–2070 (1981).

- 96 Wulf A and Nagel CW, High-pressure liquid chromatographic separation of anthocyanins of Vitis vinifera. Am J Enol Vitic 29:42-49 (1978).
- 97 Baldi A, Romani A, Mulinacci N, Vincieri FF and Casetta B, HPLC/MS application to anthocyanins of Vitis vinifera L. J Agric Food Chem 43:2104–2109 (1995).
- 98 Bartolomé B, Fernández de Simon B, Hernández T and Estrella I, Phenolic compounds in dietary fibre fractions from foods, in XVIe Journées Internationales Groupe Polyphénols and The Royal Society of Chemistry, Lisbon. 2, DTA, Lisbon, Portugal, pp 74–77 (1992).
- 99 Brandl W and Herrmann K, Hydroxyzimtsäureester der Kohlarten und der Gartenkresse. Z Lebensm Unters Forsch 176:444–447 (1983). 101 Winter M, Brandl W and Herrmann K, Bestimmung von Hydroxyzimtsäure-Derivaten in Gemüse. Z Lebensm Unters Forsch 184:11–16 (1987).
- 100 Winter M, Brandl W and Herrmann K, Bestimmung von Hydroxyzimtsäure-Derivaten in Gemüse. Z Lebensm Unters Forsch 184:11-16 (1987).
- 101 Plumb GW, Proce KR, Rhodes MJC and Williamson G, Antioxidant properties of the major polyphenolic compounds in broccoli. *Free Radic Res* 27:429–435 (1997).
- 102 Brandl W, Herrmann K and Grootjahn L, Hydroxycinnamoyl esters of malic acid in small radish (*Raphanus sativa L var sativa*). Z Naturforsch **39c**:515-520 (1984).
- 103 Tadera K and Mitsuda H, Isolation and chemical structure of a new fluorescent compound in spinach leaves. Agric Biol Chem 35:1431-1435 (1971).
- 104 Winter M and Herrmann K, Esters and glucosides of hydroxycinnamic acids in vegetables. J Agric Food Chem 34:616-620 (1986).
- 105 Williams CA, Goldstone F and Greenham J, Flavonoids, cinnamic acids and coumarins from the different tissues and medicinal preparations of *Taraxacum officinale*. *Phytochemistry* 42:121-127 (1996).
- 106 Haffke H and Engelhardt UH, Chlorogensäuren in Kaffee-Ersatzstoffen. Z Lebensm Unters Forsch 183:45–46 (1986).
- 107 Clifford MN, Shutler S, Thomas GA and Ohiokpehai O, The chlorogenic acids content of coffee substitutes. *Food Chem* 24:99–107 (1987).
- 108 Nichiforesco E, Sur la composition des derives cafeylquiniques des feuilles d/artichaut (Cynara scolymus). Plantes médicinales et Phytothérapie 4:56-62 (1979).
- 109 Horman I, Badoud R and Ammann W, Food-related applications of one- and two-dimensional high-resolution proton nuclear magnetic resonance: structure and conformation of cynarin. J Agric Food Chem 32:538–540 (1984).
- 110 Ben Hod G, Basnizki Y, Zohary D and Mayer AM, Cynarin and chlorogenic acid content in germinating seeds of globe artichoke (Cyanara scolymus L). J Gen Breed 46:63-68 (1992).
- 111 Lattanzio V and Morone I, Variations of the orthodiphenol content of *Cynara scolymus* L. during the plant growing season. *Experientia* **35**:993–994 (1979).
- 112 Martino V, Caula S, Villar S, Ferrar G and Coussio J, Compeustos polifenolicos aislados de Achyrocline tomentosa RUESBY (COMPUESTAS). Revista Latinoamericana de Quimica 19:111-113 (1988).
- 113 Reschke A, Capillargaschromatographische Bestimmung der Rosmarinsäure in Blattgewürzen. Z Lebensm Unters Forsch 176:116-119 (1983).
- 114 Malmberg A and Theander O, Free and conjugated phenolic acids and aldehydes in potato tubers. Swedish J Agric Res 14:119–125 (1984).
- 115 Malmberg AG and Theander O, Determination of chlorogenic acid in potato tubers. J Agric Food Chem 33:549–551 (1985).
- 116 Brandl W and Herrmann K, On the occurrence of chlorogenic acids in potatoes. Z Lebensm Unters Forsch 178:192– 194 (1984).
- 117 Griffiths DW, Bain H and Dale MFB, Photo-induced

changes in the total chlorogenic acid content of potato (Solanum tuberosum) tubers. J Sci Food Agric 68:105-110 (1995).

- 118 Ramamurthy MS, Maiti B, Thomas P and Nair PM, Highperformance liquid chromatography determination of phenolic acids in potato tubers (*Solanum tuberosum*) during wound healing. *J Agric Food Chem* 40:569–572 (1992).
- 119 Lewis CE, Walker JRL, Lancaster JE and Sutton KH, Determination of anthocyanins, flavonoids and phenolic acids in potatoes. 1: Coloured cultivars of Solanum tuberosum L. J Sci Food Agric 77:45-57 (1998).
- 120 Onyencho SN and Hettiarachchy NS, Antioxidant activity, fatty acids and phenolic acids composition of potato peels. J Sci Food Agric 62:345–350 (1993).
- 121 Calva-Calva G, Narbad A, Eagles J, Parr AJ, Rhodes MJC, Walton NJ, Amiot MJ and Robins RJ, Phenolic transformations by *Capsicum* spp, in *Current Trends in Fruit and Vegetables Phytochemistry*, Ed by García-Viguera C, Castañer M, Gil MI, Ferreres F and Tomás-Barberán FA, CSIC, Madrid, Spain, pp 205-209 (1995).
- 122 Elliger CA, Lundin RE and Haddon WF, Caffeoyl esters of glucaric acid in Lycopersicon esculentum leaves. Phytochemistry 20:1133-1134 (1981).
- 123 Brandl W, Galensa R and Herrmann K, HPLC analysis of celery components (hydroxycinnamic acid esters, sugars, mannitol, phthalides). Z Lebensm Unters Forsch 177:325– 327 (1983).
- 124 Dirks U and Herrmann K, 4-(β-D-glucopyranosyloxy)benzoic acid, a characteristic phenolic constituent of the Apiaceae. *Phytochem* 23:1811–1812 (1984).
- 125 Dirks U and Herrmann K, Hochleistungsflüssigkeitschromatographie der Hydroxycinnamoylchinäsauren und der 4-(β-D-Glucopyranosyloxy)-benzoesäure in Gewürzen. Lebensm Unters Forsch 179:12-16 (1984).
- 126 Scarpati ML and Oriente G, Isolamento e constituzione dell'acido rosmarinico (dal rosmarinus off). *Ricerca Sci* 28:2329-2333 (1958).
- 127 Baritaux O, Amiot MJ, Richard H and Nicolas J, Enzymatic browning of basil (*Ocimum basilicum* L). Studies on phenolic compounds and polyphenol oxidase. *Sci Aliment* 11:49– 62 (1991).
- 128 Agata L, Kusakabe H, Nishibe S, Hatano T, Yoshida T and Okuda T, Polyphenols of *Melissa officinalis*, in *Polyphenols* 96. XVIIIe Journeés Internationales Groupe Polyphénols. 1, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF. INRA Editions, Paris, France, pp 133–134 (1996).
- 129 Mendez J and Lojo MI, Phenolic and indole constituents of edible peas. J Food Sci 36:871-872 (1971).
- 130 Ferreres F, Esteban E, Carpena Ruiz R, Jimenez MA and Tomas Barberan FA, Acylated flavonol sophorotriosides from pea shoots. *Phytochemistry* **39**:1443–1446 (1995).
- 131 Strack D, Hartfeld F, Austenfeld FA, Grotjahn L and Wray V, Coumaroyl-, caffeoyl- and feruloyltartronates and their accumulation in Mung bean. *Phytochemistry* 24:147–150 (1985).
- 132 Walter WM, Purcell AE and McCollum GK, Use of highpressure liquid chromatography for analysis of sweet potato phenolics. J Agric Food Chem 27:938-941 (1979).
- 133 Kawanishi K, Yasufuku J, Ishikawa A and Hashimoto Y, Long-chain alkyl ferulates in three varieties of *Ipomoea* batatas (L.) Lam. J Agric Food Chem 38:105-108 (1990).
- 134 Goda Y, Shimizu T, Kato Y, Nakamura M, Maitani T, Yamada T, Terahara N and Yamaguchi M, Two acylated anthocyanins from purple sweet potato. *Phytochemistry* 44:183–186 (1997).
- 135 Yoshida T, Kondo T and Goto T, Unusually stable monoacylated anthocyanin from purple yam *Dioscorea alata*. *Tetrahedron Lett* **32**:5579–5580 (1991).
- 136 Cleays M, Li QM, van den Heuvel H and Dillen L, Mass spectrometric studies on flavonoid glycosides, in *Applica*tions of Modern Mass Spectrometry in Plant Science Research. Ed by Newton RP and Walton TJ, Clarendon

Press, Oxford, UK, pp 182-194 (1996).

- 137 Kroon PA, Faulds CB, Ryden P, Robertson JA and Williamson G, Release of covalently bound ferulic acid from fiber in the human colon. *J Agric Food Chem* 45:661–667 (1996).
- 138 Waldron K, Parr AJ and Ng A, Diode array HPLC analysis of dimeric cell wall phenolics, in *Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols 1*, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF. INRA Editions, Paris, France, pp 135–136 (1996).
- 139 Shibuya N, Phenolic acids and their carbohydrate esters in rice endosperm cell walls. *Phytochemistry* 23:2233–2237 (1984).
- 140 Durkee AB and Thivierge PA, Ferulic acid and other phenolics in oat seeds (Avena sativa L var Hinoat). J Food Sci 42:551-552 (1977).
- 141 Huang HM, Johanning GL and O'Dell BL, Phenolic acid content of food plants and possible nutritional implications. *J Agric Food Chem* 34:48–51 (1986).
- 142 Kroon PA, Faulds CB, Ryden P and Williamson G, Solubilisation of ferulic acid from plant cell wall materials in a model human gut system. *Biochem Soc Trans* 24:384S (1996).
- 143 Ozawa T, Nishikiori T and Takino Y, Three new substituted cinnamoylhydroxycitric acids from corn plant. Agric Biol Chem 41:359–367 (1977).
- 144 Parr AJ, Ng A, Parker M and Waldron K, Cell wall-bound phenolics of Chinese water chestnut (*Eleocharis dulcis*, Cyperaceae), in *Polyphenols 96. XVIIIe Journeés Internationales Groupe Polyphénols 2*, Ed by Vercauteren J, Chèze C, Dumon MC and Weber JF, INRA Editions, Paris, France, pp 321-322 (1996).
- 145 Buchanan CJ, Wallace G, Fry SC and Eastwood MA, In vivo release of ¹⁴C-labelled phenolic groups from intact dietary spinach cell walls during passage through the rat intestine. *J Sci Food Agric* 71:459–469 (1996).
- 146 Seitz LM, Stanol and sterol esters of ferulic and p-coumaric acids in wheat, corn, rye, and triticale. J Agric Food Chem 37:662-667 (1989).
- 147 Norton RA, Isolation and identification of steryl cinnamic acid derivatives from corn bran. *Cereal Chem* 71:111–117 (1994).
- 148 Strack D, Engel U, Weissenbôck G, Grotjahn L and Wray V, Ferulic acid esters of sugar carboxylic acids from primary leaves of rye (Secale cereale). Phytochemistry 25:2605-2608 (1986).
- 149 Daniels DGH, King HGC and Martin HF, Antioxidants in oats: esters of phenolic acids. J Sci Food Agric 14:385–390 (1963).
- 150 Daniels DGH and Martin HF, Isolation of a new antioxidant from oats. *Nature* **191**:1302 (1961).
- 151 Daniels DGH and Martin HF, Structures of two antioxidants isolated from oats. *Chem Ind* 2058 (1964).
- 152 Daniels DGH and Martin HF, Antioxidants in oats: monoesters of caffeic and ferulic acids. J Sci Food Agric 18:589– 597 (1967).
- 153 Daniels DGH and Martin HF, Antioxidants in oats: glyceryl esters of caffeic and ferulic acid. J Sci Food Agric 19:710– 712 (1968).
- 154 Collins FW, Oat phenolics: avenanthramides, novel substituted N-cinnamoylanthranilate alkaloids from oat groats and hulls. J Agric Food Chem 37:60-66 (1989).
- 155 Collins FW, McLachlan DC and Blackwell BA, Oat phenolics: avenalumic acids, a new group of bound phenolic acids from oat groats and hulls. *Cereal Chem* 68:184–189 (1991).
- 156 Dimberg LH, Theander O and Lingnert H, Avenanthramides – a group of phenolic antioxidants in oats. Cereal Chem 70:637-641 (1993).
- 157 McMurrough I, Roche GP and Cleary KG, Phenolic acids in beers and worts. *J Inst Brew* 90:181–187 (1984).
- 158 Hollman PC, van Trijp JM, Mengelers MJ, de Vries JH and Katan MB, Bioavailability of the dietary antioxidant flavonol quercetin in man. *Cancer Lett* **114**:139–140 (1997).